SHRI SHIVAJI EDUCATION SOCIETY AMRAVATI'S SCIENCE COLLEGE, CONGRESS NAGAR, NAGPUR

PG Department of Microbiology Add on Course: CRISPR Cas9 Genome Editing Session 2022-23

Course Coordinator Report

A free Add-On Course for PG students in the Department Microbiology, Shri Shivaji Education Society Amravati's Science College, Congress Nagar, Nagpur was held from 01st April 2024 to 17th April 2024. The course title was "CRISPR Cas9 Genome Editing". It is the complete beginner to Expert Course was perfect for anyone who wants to learn CRISPR Cas9 Genome Editing.

The CRISPR Cas9 Genome Editing course is designed to provide a comprehensive overview of CRISPR-Cas9 technology, its mechanisms, and its applications in gene editing. It includes both theoretical knowledge and practical experience, equipping students with the skills needed for genome editing and molecular biology techniques.

The course duration was 10 weeks (30 hours). Two theory classes were engaged on Friday & Saturday and one Practical was engaged in every week. The structure of marking system was 50 marks on theory paper and 40 marks on practical examination including 10 marks for internal. The question paper of theory examination was in MCQ type of 25 questions with four multiple choices. Practical examination was also taken on this course for 40 marks. Internal marks assessment was on the basis of regularity, attendance, assignment submission etc. All the 30 students were present in both theory and practical examination. The result was prepared and certificates were also distributed to the students.

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PG Department of Microbiology Add on Course: CRISPR-Cas9 Genome Editing Session 23-24

The Principal SSES Ami's Science College. Congress Nagar, Nagpur-12

Subject: For permission to conduct the add on courses in Microbiology and Biotechnology department during the session 2023-2024

Respected Sir.

This is to request you that, the teachers of Microbiology and Biotechnology department have prepared the syllabus and modules of the 30 hours certificate courses for the session 2023-2024.

The details of the course module, syllabus and time table is submitted here with.

Hence please permit to run the add on courses and obline me

Thanking you

Yours sincerely

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Permitted per

SHRI SHIVAJI EDUCATION SOCIETY AMRAVATI'S SCIENCE COLLEGE, CONGRESS NAGAR, NAGPUR

PG Department of Microbiology NOTICE

Date: 11/03/2024

All the students are informed that P.G. Department of Microbiology runs Add on Course: CRISPR-Cas9 Genome Editing for the session 2023-24. Interested students of M.Sc. are requested to register their names to the course Coordinator Dr. Pranita B. Gulhane on or before 29/03/2024.









U.G. DEPARTMENT OF MICROBIOLOGY, SCIENCE COLLEGE, CONGRESS NAGAR, NAGPUR

Accredited with CGPA of 3.51 at 'A+' Grade by NAAC. Bangalore
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Recognized Center for Higher Learning & Research
A Mentor College under Paramarsh Scheme of UGC. New Delhi
A Mentor College under Paris Sparsh Scheme of Maharashtra State

Add on Course for the Session 2023-24 CRISPR-Cas9 Genome Editing

Course Introduction

This course provides a comprehensive overview of CRISPR-Cas9 technology, its mechanisms, and its applications in gene editing. It includes both theoretical knowledge and practical experience, equipping students with the skills needed for genome editing and molecular biology techniques.

Course Objectives

- Understand the fundamentals of CRISPR-Cas9 technology and its historical development.
- Learn the molecular mechanisms of CRISPR-Cas9 and its components.
- Explore the various applications of CRISPR-Cas9 in gene editing across different organisms.
- Gain hands-on experience in designing and executing CRISPR experiments, including data analysis and troubleshooting.

Registration Date: 29/03/2024

Prof. Atul Bobdey
Coordinator
Dept. of Microbiology

Prof. Mahendra Dhore
Principal
Science College, Nagpur

PG Department of Microbiology

Add on Course: Syllabus for CRISPR-Cas9 Genome Editing (Session 2023-24)

Course Co-ordinator: Dr. Pranita B. Gulhane

Course Introduction

This course provides a comprehensive overview of CRISPR-Cas9 technology, its mechanisms, and its applications in gene editing. It includes both theoretical knowledge and practical experience, equipping students with the skills needed for genome editing and molecular biology techniques.

Course Objectives

- Understand the fundamentals of CRISPR-Cas9 technology and its historical development.
- Learn the molecular mechanisms of CRISPR-Cas9 and its components.
- Explore the various applications of CRISPR-Cas9 in gene editing across different organisms.
- Gain hands-on experience in designing and executing CRISPR experiments, including data analysis and troubleshooting.

Instructional Strategies: Theory class, Practical, Video clips, Models etc.

Evaluation Strategies: Oral discussions and Final MCQ examination

Course Outcomes

Upon completing this course, students will be able to:

- Describe the principles and components of the CRISPR-Cas9 system.
- Design CRISPR-Cas9 constructs for specific gene targets.
- Perform CRISPR-Cas9 mediated gene editing experiments.
- Analyze and interpret the results of CRISPR-Cas9 experiments.
- Discuss the ethical and societal implications of genome editing technology.

Duration of course: Ten weeks (30 Hours)

Module: The Structure of Syllabus and system of evaluation

		Total Marks		
Course	Theory Papers and Practical	Theory	Internal	Practical
Certificate Course in CRISPR-Cas9	Theory paper- CRISPR-Cas9 technology * Theory examination will be of MCQ pattern having 25 questions each with equal marks.	50	10	40
technology	* Practical examination will be based on performance evaluation in the laboratory and hands-on-training	100		

Dr. Pranita Gulhane Add on Course Coordinator

Dr. Amitabh Halder

Prof. Mahendra Dhore

Principal

Buore

IQAC Coordinator Internal Quality Assurance Cell (IQAC)

Principal S. S. E. S. Amravati's

S. S. E. S. A. Science College Science College, Nagpur. Congress Nagar, Nagpur.



PG Department of Microbiology Syllabus of Add on Course: Syllabus for CRISPR-Cas9 Genome Editing (Session 2023-24)

Course Units

Unit 1: Introduction to CRISPR-Cas9

- · History and Discovery of CRISPR-Cas Systems
- Mechanisms of CRISPR-Cas9
- Structure and function of Cas9
- Guide RNA (gRNA) design
- · Types of CRISPR Systems
- Class 1 and Class 2 CRISPR systems
- · Applications and differences

Unit 2: Designing CRISPR-Cas9 Experiments

- Target Selection
- · Criteria for selecting target sites
- · Off-target effects and their minimization
- gRNA Design and Synthesis
- · Tools and software for gRNA design
- · Synthesis and validation of gRNAs
- CRISPR Vectors and Delivery Methods
- Plasmid vectors, viral vectors, and ribonucleoprotein complexes
- Methods of delivery (transfection, electroporation, etc.)

Unit 3: Applications of CRISPR-Cas9

- Gene Knockout and Gene Knock-in
- · Techniques for generating knockouts and knock-ins
- · Applications in model organisms and cell lines
- CRISPR in Disease Models and Therapeutics
- · CRISPR for disease research
- · Therapeutic applications and clinical trials
- · CRISPR Beyond Gene Editing

- CRISPRa and CRISPRi for gene regulation
- · Base editing and prime editing

Unit 4: Data Analysis and Ethical Considerations

- · Data Analysis and Validation
- · Methods for detecting and analyzing CRISPR edits
- · Sequencing and bioinformatics tools
- · Troubleshooting CRISPR Experiments
- · Common issues and solutions
- · Ethical, Legal, and Social Implications
- · Ethical considerations in genome editing
- · Regulatory landscape and public perception

Practical Sessions

Practical 1: Designing gRNAs,

- · Use of bioinformatics tools to design gRNAs for a given target gene.
- Synthesis and validation of gRNAs.

Practical 2: Cloning and Delivery of CRISPR-Cas9 Constructs

- Cloning gRNA into CRISPR vectors.
- · Transfection of CRISPR constructs into cells.

Practical 3: Gene Editing and Screening

- · Performing CRISPR-Cas9 mediated gene editing.
- Screening and validating edited cells using PCR and sequencing.

Practical 4: Data Analysis and Troubleshooting

- Analyzing sequencing data to confirm gene edits.
- Troubleshooting common issues in CRISPR experiments.

Dr. Pranita B. Gulhane Course- Coordinator

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Add on Course

Week-wise teaching plan:

Week	Hrs.	Syllabus			
Week 1	1	Structure and function of Cas9			
	1	Guide RNA (gRNA) design			
	1	Class 1 and Class 2 CRISPR systems			
Week 2	1	Criteria for selecting target sites			
	1	Off-target effects and their minimization			
	1 .	Tools and software for gRNA design			
Week 3	1	Synthesis and validation of gRNAs			
	1	Plasmid vectors, viral vectors, and ribonucleoprotein complexes			
	1	Methods of delivery (transfection, electroporation, etc.)			
Week 4	1	Techniques for generating knockouts and knock-ins			
	1	Applications in model organisms and cell lines			
	1	CRISPR for disease research			
Week 5	1	Therapeutic applications and clinical trials			
	2	CRISPRa and CRISPRi for gene regulation			
Week 6	2	Base editing and prime editing			
	1	Methods for detecting and analyzing CRISPR edits			

Week 7	2	Sequencing and bioinformatics tools
	2	Common issues and solutions
Week 8	2	Ethical considerations in genome editing
	2	Regulatory landscape and public perception
Week 9	1	Practical 1: Designing gRNAs
	1	Practical 2: Cloning and Delivery of CRISPR-Cas9 Constructs
Week 10	1	Practical 3: Gene Editing and Screening
	1	Practical 4: Data Analysis and Troubleshooting



Time Table

w.ef. 01/04/2024

Day	Theory				
Friday	Dr. Pranita Gulhane (R. no C6) Theory 4.00 PM - 5.00 PM				
Saturday	Dr. Pranita Gulhane (R. no C6) Practical 4.00 PM - 5.00 PM				
	Dr. Pranita Gulhane (R. no C6) Theory 4.00 PM - 5.00 PM				

SEAL SEAL STORY

Dr. Pranita B. Gulhane Course- Coordinator

Add on Course

SHRI SHIVAJI EDUCATION SOCIETY AMRAVATI'S SCIENCE COLLEGE, CONGRESS NAGAR, NAGPUR

PG Department of Microbiology

EXAMINATION NOTICE

Date: 22/04/2024

All the students enrolled for **Add on Course: CRISPR-Cas9 Genome Editing** for the session 2023-24 are informed that Theory and Practical Exam of the course is scheduled on 26/04/2024. All the appearing students are informed to remain present in Microbiology Laboratory at 10:30 – 11:30AM AM for Theory Exam and at 12:30PM – 5:30PM for Practical Exam.

SEAL SEAL STATES

Dr. Pranita B. Gulhane Course- Coordinator Add on Course

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List of the Students: Add on Course- CRISPR-Cas9 Genome Editing (Session 2023-2024)

Sr. No.	Name of Student	Signature
1)	Akanksha R. Bisen	Bisen.
2)	Akanksha V. Tekade	Stehade
3)	Anisha A. Shende	-A Shonde
4)	Anuradha P. Khope	Spape
5)	Arshiya S. Mushtaque	Amushdagu
6)	Arya S. Walode	A Walades
7)	Ashwini D. Nandanwar	Afterndenna
8)	Asmita A. Wagh	weagh
9)	Astha A. Sakharwade	Ashar
10)	Bhisvani M. Dhurve	Bahune
11)	Diksha S. Choube	- 16houha
12)	Ishita Y. Padgil	Ishita
13)	Ishwari N. Gawande	I Cresson do
14)	Janhavi D. Parbat	Togobat
15)	Leena N. Meher	Acha!
16)	Manisha G. Lilhare	Lithour
17)	Manisha R. Roy	heay
18)	Namrata O. Nagose	W. Nagose
19)	Ojaswini R. Bhagat	Oplanda
20)	Rujuta R. Ramteke	Divi
21)	Rutuja R. Meshram	d'anteker

22)	Sejal U. Nilatkar	Milather
23)	Shamim M. Shahzad	Shazust
24)	Shivani R. Vikhar	cy khan
25)	Simran V. Bode	Stubolo
26)	Snehal N. Sahare	Lahase.
27)	Sushma D. Lilhare	Stiller
28)	Vaibhav S. Jaronde	Varonde
29)	Vami K. Masram	Morrans
30)	Yashoda R. Wade	lastique



Dr. Franta B. Gullare

Theory Exam Multiple Choice Questions (MCQs) Pattern

- 1. What does CRISPR stand for?
- A) Clustered Regularly Interspaced Short Palindromic Repeats
- B) Clustered Repeatedly Interspaced Short Palindromic Repeats
- C) Controlled Regularly Interspaced Short Palindromic Repeats
- D) Clustered Regularly Interspaced Short Polymeric Repeats
- Answer: A
- 2. Which enzyme is primarily used in the CRISPR-Cas9 system for cutting DNA?
- A) Cas1
- B) Cas2
- C) Cas9
- D) Cas12
- Answer: C
- 3. What is the role of the guide RNA (gRNA) in CRISPR-Cas9?
- A) To cut the DNA
- · B) To bind to the target DNA sequence
- · C) To repair the DNA break
- · D) To synthesize new DNA
- · Answer: B
- 4. Which of the following is NOT a method for delivering CRISPR-Cas9 into cells?
- A) Transfection
- B) Electroporation
- C) Transduction
- D) PCR
- Answer: D
- 5. What is the main advantage of using CRISPR-Cas9 over previous genome editing technologies?
- A) Higher specificity
- B) Lower cost
- · C) Easier design and implementation
- · D) All of the above
- · Answer: D
- 6. Off-target effects in CRISPR-Cas9 are due to:
- A) Inaccurate DNA cutting
- B) Mismatches between gRNA and non-target DNA
- C) Incomplete guide RNA
- · D) None of the above
- Answer: B
- 7. Which of the following CRISPR systems is classified as a Class 2 system?
- A) Type I
- B) Type II
- · C) Type III

- D) Type IV
- · Answer: B
- 8. In CRISPR-Cas9 technology, what does PAM stand for?
- · A) Protospacer Adjacent Motif
- B) Protospacer Additional Motif
- · C) Protospacer Aligned Motif
- · D) Protospacer Altered Motif
- Answer: A
- 9. CRISPR-Cas9 can be used to:
- A) Knockout genes
- · B) Insert genes
- · C) Edit specific DNA sequences
- D) All of the above
- Answer: D
- 10. Which tool is commonly used for designing gRNAs?
- A) BLAST
- B) ClustalW
- C) CRISPRdirect
- D) ExPASy
- Answer: C
- 11. What is a potential application of CRISPR-Cas9 in agriculture?
- · A) Developing disease-resistant crops
- . B) Increasing crop yield
- · C) Enhancing nutritional content
- D) All of the above
- Answer: D
- 12. Which component of the CRISPR-Cas9 system is responsible for recognizing the target DNA sequence?
- A) Cas9 enzyme
- B) Donor DNA
- C) Guide RNA (gRNA)
- D) PAM sequence
- Answer: C
- 13. What type of gene editing involves introducing a new gene at a specific location in the genome?
- · A) Gene knockout
- · B) Gene knock-in
- · C) Gene silencing
- D) Gene duplication
- Answer: B
- 14. CRISPRa and CRISPRi are used for:
- · A) Activating or inhibiting gene expression
- . B) Cutting DNA
- · C) Repairing DNA
- D) Sequencing DNA
- Answer: A
- 15. Which technique can be used to detect CRISPR-Cas9 induced mutations?
- · A) Western blotting

- B) PCR
- C) Gel electrophoresis
- D) Sanger sequencing
- Answer: D
- 16. Which application of CRISPR-Cas9 has entered clinical trials for treating genetic disorders?
- A) Gene knockout
- B) Gene editing in embryos
- C) Gene therapy for inherited diseases
- D) All of the above
- Answer: C

17. In CRISPR-Cas9, the Cas9 enzyme creates a:

- A) Single-strand break
- B) Double-strand break
- C) Triple-strand break
- D) None of the above
- Answer: B

18. What is the purpose of the repair template in CRISPR-Cas9 gene editing?

- A) To guide the Cas9 enzyme
- B) To provide a sequence for homologous recombination
- C) To cut the DNA
- D) To bind to the target DNA
- Answer: B

19. What ethical concern is associated with CRISPR-Cas9 technology?

- A) Off-target effects
- B) Germline editing
- C) Gene drives in wild populations
- D) All of the above
- Answer: D

20. Which of the following is a limitation of CRISPR-Cas9?

- A) High specificity
- B) Low cost
- C) Potential for off-target effects
- D) Ease of use
- Answer: C

21. Which alternative to CRISPR-Cas9 is known for introducing precise point mutations without double-strand breaks?

- A) TALENs
- B) ZFNs
- C) Base editors
 - D) Meganucleses
 - Answer: C

22. Which of the following is a common method to validate CRISPR-Cas9 edits?

- A) RNA sequencing
- B) Chromatography
- C) Western blot
- D) T7E1 assay
- Answer: D

23. The specificity of CRISPR-Cas9 can be increased by:

- · A) Using a shorter gRNA
- . B) Increasing Cas9 concentration
- . C) Designing gRNA with fewer mismatches
- D) Using a high-fidelity Cas9 variant
- Answer: D
- 24. What is the primary advantage of using CRISPR-Cas9 over traditional breeding methods in agriculture?
- . A) Faster generation of desired traits
- . B) More cost-effective
- · C) More precise changes
- . D) All of the above
- · Answer: D
- 25. Which regulatory body oversees the approval of CRISPR-based therapies in the United States?
- A) EPA
- B) FDA
- C) USDA
- D) NIH
- · Answer: B

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Practical Exam Question Paper:

Subject

: CRISPR-Cas9 Genome Editing

Center

: S.S.E.S.A's Science College, Nagpur

Time

: 5 hrs per day

Dates

: 26/04/2024

Max. Marks: 40

Q.1. Cloning and Delivery of CRISPR-Cas9 Constructs	10
	- 7/2
Q.2. Gene Editing and Screening	10
Q.3. Viva-Voce	10
Q.4. Practical Record	10

Total Marks 40

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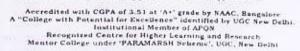
OMR Answer Sheet



Shri Shivaji Education Society, Amravati's

SCIENCE COLLEGE

Congress Nagar, Nagpur-12 (M.S.), India





P.G. DEPARTMENT OF MICROBIOLOGY

Add-on Course Course Exam Name: Add on Course in CRISPR-Cas 9 Genome Editing Name of Student: INSTRUCTIONS FOR FILLING THE SHEET A Kanksha Bisan 2023-24

2 Use only blue black ball sown 2 Use of pencil is strictly prohibited 3 Use of pencil is strictly prohibited 4 Circles should be darkened completely and properly 4 Circles should be darkened completely and properly This sheet should not be folded or crushed Cutting and erasing on this sheet is not allowed.
 Do not use any stray marks on the sheet. Test Date: 22/04/2024 Max. Marks: 50 Do not use marker or white fluid to hide the mark WADNG METHODS CORRECT METHOD 0000 48 Obtained Marks: Invigilator Signature 41 0 0 0 0 ABCD ABCD ABCD ABCD 110000 110000 21 00000 1 .000 42 0000 32 0000 22 () () () 12 0000 20000 41 0000 11 0000 23 0000 13 (6 0 0 30000 44 0000 34 00000 24 0 0 0 9 14 . 000 40000 45 0000 35 0000 25 00 00 15 0000 50000 46 0000 36 0000 26 0000 16 00 60 6000 47 0000 37 0000 27 0000 17 0 000 10000 48 0000 38 0000 28 0000 18 0 0 0 0 49 0000 39 0000 29 0000 19 0000 90000 50 0000 400000 30 0000 20 00 00 10 0000

Sr. No.	Name of Student	Marks obtained out of 50 (Theory)	Marks obtained out of 40 (Practical)	Marks obtained out of 10 (Internal)	Total Marks 100	Grade
1)	Akanksha R. Bisen	48	36	10	94	0
2)	Akanksha V. Tekade	42	35	10	87	A+
3)	Anisha A. Shende	48	36	10	94	0
4)	Anuradha P. Khope	44	35	10	89	A+ *
5)	Arshiya S. Mushtaque	50	34	10	94	0
6)	Arya S. Walode	48	35	10	93	0
	Ashwini D.					
7)	Nandanwar	42	34	10	86	A+
8)	Asmita A. Wagh	48	35	10	93	0
9)	Astha A. Sakharwade	50	37	10	97	0
10)	Bhisvani M. Dhurve	42	34	10	86	A+
11)	Diksha S. Choube	44	35	10	89	A+
12)	Ishita Y. Padgil	48	36	10	94	0

13)	Ishwari N. Gawande	50	38	10	98	0
14)	Janhavi D. Parbat	48	35	10	93	0
15)	Leena N. Meher	46	39	10	95	0
16)	Manisha G. Lilhare	42	35	10	87	A+
17)	Manisha R. Roy	44	35	10	89	A+
18)	Namrata O. Nagose	48	35	10	93	0
19)	Ojaswini R. Bhagat	42	35	10	87	A+
20)	Rujuta R. Ramteke	50	38	10	98	0
21)	Rutuja R. Meshram	50,	34	10	94	0
22)	Sejal U. Nilatkar	42	35	10	87	A+
23)	Shamim M. Shahzad	44	35	10	89	A+
24)	Shivani R. Vikhar	42	34	10	86	A+
25)	Simran V. Bode	50	38	10	98	0
26)	Snehal N. Sahare	48	35	10	93	0
27)	Sushma D. Lilhare	48	35	10	93	0
28)	Vaibhav S. Jaronde	50	38	10	98	0
29)	Vami K. Masram	50	34	10	94	0
30)	Yashoda R. Wade	48	36	10	94	0





Shri Shivaji Education Society Amravati's

SCIENCE COLLEGE, CONGRESS NAGAR,

NAGPUR

Accredited with CGPA of 3.51 at 'A+' Grade A College with Potential for Excellence



CERTIFICATE

Mr./Ku. Lkanksha R. Bisen

is awarded with certificate on successful completion of the

course entitled, Certificate course in "CRISPR-Cash 9 Gename Editing"

Session 2023-24 under Add-on course conducted for 30 hours from 15/12/2023 to 24/02/2024 by Department of Microbiology, SSESA's, Science College, congress Nagar, Nagpur 440012.

He/She has passed the Examination with '0' Grade.



Dr. Pranita Gulhane

Coordinator, Department of Microbiology



Prof. M. P. Dhore

Principal, Science College, Nagpur